## **The Detection of Designer Drugs from Plasma** пп Greta J. Ren and **IUPUI** Department of Chemistry and Chemical Biology, **SCHOOL OF SCIENCE**

## **Overview**

- **Designer drugs are not detected by routine drug screens, and are more potent than traditional drugs**
- A disposable paper spray cartridge with SPE column can carry out analyte pre-concentration and ionization
- Method optimized for detection of two synthetic cannabinoids JWH-200 and JWH-250
- Most frequently abuse synthetic cannabinoids can be detected at sub-ng/mL levels

## **Introduction**

- Designer drugs mimic psychoactive effects of traditional drugs, however, they are typically more potent and can have unpredictable and severe health effects
- They are cheap and marketed as 'legal highs', since they cannot be detected by routine drug screens
- New (often more dangerous) drugs continue to emerge as known designer drugs become banned
- There is a need for a rapid and sensitive analytical method to detect designer drugs





**Figure 1: Structure of THC, and synthetic cannabinoids JWH-250 and 5F-ADB Figure 2: Cartridge Cartridge in the Structure 2: Cartridge** 

**1.** Sample is loaded at the top of the SPE column, and allowed to wick through idge to help remove matrix components

MS inlet and spray solvent is added to the top to extract

analyte signal is collected (2-5 minutes)



**Extraction and drying ipped with SPE<sup>1</sup>** 

- Paper spray mass spectrometry can directly analyze biological samples
- Advantages: no sample preparation, small sample volume, small solvent volume, no solvent waste, no carry over, rapid analysis (1-2 minute run), automatable
- Cartridge equipped with solid phase extraction (SPE) column can perform analyte pre-concentration and ionization
- SPE helps improve detection limits by allowing larger sample volumes to be used, removing matrix interferences and pre-concentrating the analytes

**positioned in front the of the mass spectrometer inlet for analysis**

## **Methods**





**Elution and detection** 

Cannabinoid	<b>ISTD</b>	<b>Transitions</b>
<b>JWH-200</b>	AB-CHMINACA d4	$385.3 \rightarrow 155.0494$
$JWH-250$	AB-CHMINACA d4	$336.3 \rightarrow 121.0652$
AM-2201	AB-CHMINACA d4	$360.2 \rightarrow 155.0494$
<b>AB-CHMINACA</b>	AB-CHMINACA d4	$357.2 \rightarrow 312.2076$
$5F-ADB$	AB-FUBINACA d4	$378.2 \rightarrow 251.1193$
$5F-PB-22$	AB-CHMINACA d4	$385.3 \rightarrow 232.1135$
$XLR-11$	AB-CHMINACA d4	$330.3 \rightarrow 125.0966$
THJ-2201	AB-FUBINACA d4	$361.2 \rightarrow 251.1193$

- Cartridges were made from Delrin® on a milling machine
- Two parts of the cartridge join together via tongue and groove
	- Bottom part dimensions: 40mm x 26mm x 6  $mm$  (LWH)
	- Top part dimensions: 14mm x 22mm x 13mm (LWH)
- Bottom part has two separate regions to hold absorbent pad and paper spray substrate
- Top part contains the SPE column (3.0 mm Whatman ET31 paper punch, SPE material, 3.0 mm nylon punch)
- Mass spectrometry analysis was performed using Thermo Scientific TSQ Vantage (TSQ) in the multiple reaction monitoring (MRM) mode and Thermo Scientific Q-Exactive Focus (QE) in the parallel reaction monitoring (PRM) mode





**Table 1: TSQ MRM transitions, and the ISTD used for normalization \*Quantitation transition is bolded**

**Table 2: QE PRM transitions, and the ISTD used for normalization**

- QE and TSQ produced different fragmentation and different MS/MS spectra
- The fragments with the highest intensity were selected for quantitation





## **Conclusions**

## **References**

## **Acknowledgments**

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**1.** Zhang, C. & Manicke, N. E. Development of a Paper Spray Mass Spectrometry Cartridge with Integrated Solid Phase Extraction for Bioanalysis. *Anal. Chem.* **87,** 6212–6219 (2015)

2.5 2 Ratio (Analyte/ISTD) **Ratio (Analyte/ISTD)** 1.5 1 0.5 0  $\bullet$  JWH-200  $\Box$  JWH-250  $\triangle$  AM-2201  $\boxtimes$  AB-CHMINACA  $\angle$  5F-PB-22  $\angle$  XLR-11  $\angle$  THJ-2201





**Figure 12: Calibration curve for synthetic cannabinoids, MS analysis performed using TSQ**

**Figure 13: Calibration curve for synthetic cannabinoids, MS analysis performed using QE**

- Rinsing SPE column with water after loading the sample helps remove matrix components
- Washing the paper substrate helps reduce the background signal
- For 100 µL of plasma, 10 mg of SPE material gave the best results
- Signal to Noise ratio (S/N) increases with larger sample volumes
- Optimized method was used to analyze samples on QE





- All synthetic cannabinoids could be detected subng/mL levels
- Optimized SPE method decreased the detection limits  $~100$  times
- Good linearity from 0.1  $10 \text{ ng/mL}$
- Some adjustments may be necessary to achieve the same LODs with the QE

**Table 4: Limits of detection and R<sup>2</sup> obtained from synthetic cannabinoid calibration curves**

- A method was developed and optimized for synthetic cannabinoids JWH-200 and JWH-250 **Extraction solvent, SPE sorbent, sample volume, SPE amount and wash steps were investigated**
- Method was able to detect several synthetic cannabinoids that were most commonly detected in US toxicology labs in the last two years at sub-ng/mL concentrations
- Synthetic cannabinoids can be quantified with the use of an ISTD
- The presented method allows for rapid, sensitive (sub ng/mL) detection of synthetic cannabinoids

# **Racking Drugs Spectrometry Cartridge** Nicholas Manicke <sup>7</sup>, Indiana University-Purdue University Indianapolis

**Results**



- Acetonitrile with 0.1% formic
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